



Office: 336.368.4500
Mobile: 336.575.2343
jmcmanus@allsenvironmental.com

February 4, 2020

Jay Fuller
Director of Facilities and Maintenance

Alamance-Burlington School System
307 Prison Camp Road
Graham, North Carolina 27253

Subject: Report of Airborne and Surface Microbial Sampling
Eastlawn Elementary School – Room #20A
502 N. Graham-Hopedale Road
Burlington, North Carolina
Project No.: 201-2001-02

Dear Mr. Fuller:

ALIS has completed the airborne microbial sampling at Eastlawn Elementary School in Burlington, North Carolina. On January 29, 2020 we conducted air sampling for the presence of airborne fungi at the subject property. The purpose of our sampling was to determine the presence and species of airborne fungi and the degree of concentration within the area of concern. One air sampling pump was calibrated and placed in room 20A, and one sample was collected from outside the building as a reference (baseline) for comparison to the inside conditions. Additionally, we obtained a sample of suspect growth from the window sash for confirmation of species. No other areas of the building were included in the scope of work.

Results

The laboratory results found a significantly higher level of Cladosporium on the indoor air sample when compared to the outdoor air sample. Lower levels of Aspergillus/Penicillium were also detected on the indoor sample. Cladosporium is often found indoors in a high humidity environment. During the sampling period, temperature and humidity readings were taken in room 20A and outside the building. The humidity reading in room 20A was 62% while the outdoor reading was only 33%. Temperature indoor was 68°F while the outdoor reading was 52°F. I observed condensation on the window's glass panes and sills. The laboratory analysis on the surface sample obtained from the window sash identified the visible growth as Cladosporium with a moderate spore count estimate.

A Protimeter moisture indicator was used to locate moisture within the block walls. The moisture indicator detected wet conditions within the block walls in the perimeter wall below the windows and in the corner at the opposing wall. Sample results and additional information on fungal spores are attached to this report: “Spore Trap Analysis” and “Direct Analysis.”

Sampling Methodology

Non-viable samples were collected with a spore trap slide using Allergenco-D Cassettes mounted to a sampling pump. The cassettes contain glass slides that are coated with a sticky substance that captures airborne particulates that impinge on the slides. The air samples were collected at 15 liters per minute for 10 minutes. Calibration of sampling equipment was performed with a precision rotameter (a secondary calibration source). Rotameters are calibrated against a primary standard. Field calibration was performed before and after sampling. The air samples were sealed for transport to Hayes Microbial Consulting in Midlothian, Virginia for analysis. Hayes Microbial is a participant in the American Industrial Hygiene Association, Laboratory Accreditation Program (AIHA-LAP) for Environmental Microbiology.

Background Information on Mold in Buildings

Mold spores exist normally in outdoor and indoor air and can be measured in air and carpets of normal homes, office buildings, hospitals and schools. Naturally occurring sources of mold spores include soil, plants and other sources. The air concentration of these normally occurring mold spores is dependent on the season, environmental conditions and other factors. Elevated levels of mold in building materials may occur if chronic moist conditions from water leaks, floods, chronic high relative humidity, or malfunctioning heating, ventilation or air conditioning systems, allow moisture to remain for prolonged periods on organic matter in the presence of warm ambient temperatures. Under these conditions, low levels of fungal spores in air, plants or other sources, may proliferate on cellulose containing materials such as carpets, wallboard, wood, paper or dusty surfaces (which may serve as a food source), and result in mold contamination. Many fungal spores are allergenic to susceptible persons exposed, though individual susceptibility varies greatly. There is no practical way to eliminate all mold and mold spores in the indoor environment; the way to control indoor mold growth is to control moisture.

ALIS appreciates the opportunity to be of service to you on this project. We would welcome the opportunity to discuss at your convenience, any of the results contained in this report. Please contact us if you have any questions or if we may be of further service.

Sincerely,
ALIS ENVIRONMENTAL, INC.



James P. McManus
Vice-President

Attachments: “Spore Trap Analysis”
“Direct Analysis”



#20004000

Analysis Report prepared for

ALIS Environmental Inc.

1027 Koontz Haven Rd
Pinnacle, NC 27043

Phone: (336) 368-4500

201-2001-02
Eastlawn Elementary School
502 N. Graham - Hopedale Rd
Burlington, NC

Collected: **January 29, 2020**
Received: **January 31, 2020**
Reported: **January 31, 2020**



EPA Laboratory ID: VA01419

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 4 samples by FedEx in good condition for this project on January 31st, 2020.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC..

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.

Steve Hayes, BSMT(ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.



Lab ID: #188863



DPH License: #PH-0198

Sample Number	02276037	02330897	02331817			
Sample Name	Outside Bldg	Room 20A	Blank			
Sample Volume	150.00 liter	150.00 liter	0.00 liter			
Reporting Limit	7 spores/m ³	7 spores/m ³	1 spore/m ³			
Background	2	2	NBD			
Fragments	40/m ³	ND	ND			
Organism	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total
Alternaria						
Ascospores	3	20	33.3%	4	27	4.2%
Aspergillus/Penicillium				22	147	22.9%
Basidiospores						
Bipolaris/Drechslera				1	7	1.0%
Chaetomium				67	447	69.8%
Cladosporium				1	7	1.0%
Curvularia						
Epicoccum	6	40	66.7%			
Fusarium						
Memnoniella						
Myxomycetes				1	7	1.0%
Pithomyces						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Total	9	60	100%	96	642	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
Collected: Jan 29, 2020	Received: Jan 31, 2020	Reported: Jan 31, 2020		
Project Analyst: Avani Devmurari, MS	Reviewed By: Steve Hayes, BSMT	Date: 01 - 31 - 2020	Date: 01 - 31 - 2020	
3005 East Boundary Terrace, Suite F. Midlothian, VA, 23112		contact@hayesmicrobial.com		Page: 2 of 9



Jim McManus
ALIS Environmental Inc.
1027 Koontz Haven Rd
Pinnacle, NC 27043
(336) 368-4500

201-2001-02
Eastlawn Elementary School
502 N. Graham - Hopedale Rd
Burlington, NC

#20004000

Direct Analysis
SOP - HMC#102



#4	Bio-Tape (1.00 cm2)	Organism	Spore Estimate	Mycelial Estimate
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B2043579 - Room 20A Window Sash

Cladosporium

Moderate

Many

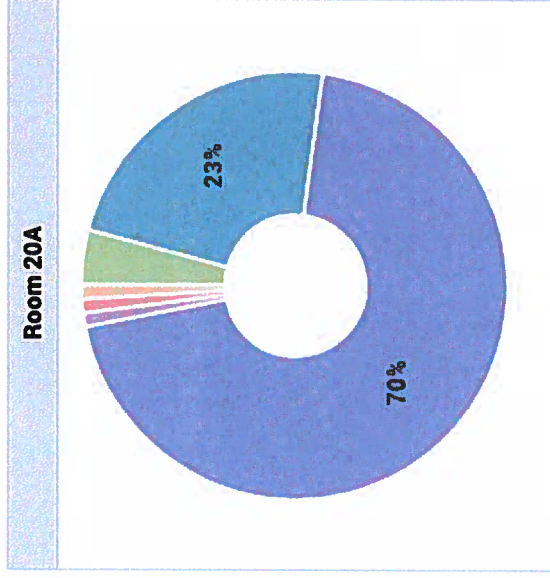
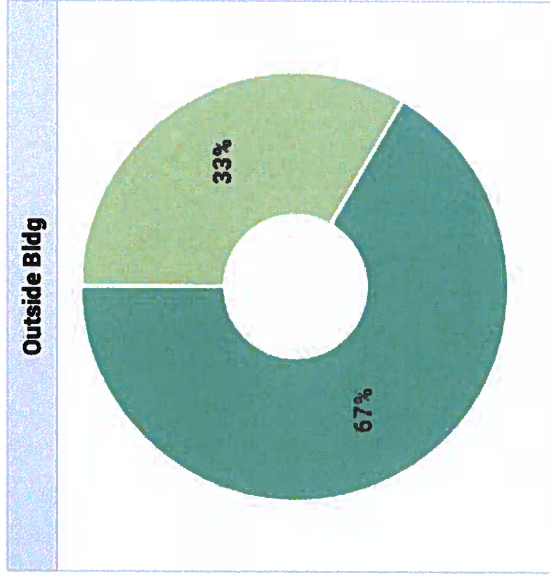
Collected: Jan 29, 2020	Received: Jan 31, 2020	Reported: Jan 31, 2020
Project Analyst: Avani Devmurari, MS	Date: 01 - 31 - 2020	Date: 01 - 31 - 2020
	Reviewed By: Steve Hayes, BSMT	
3005 East Boundary Terrace, Suite F. Midlothian, VA. 23112	(804) 562-3435	contact@hayesmicrobial.com



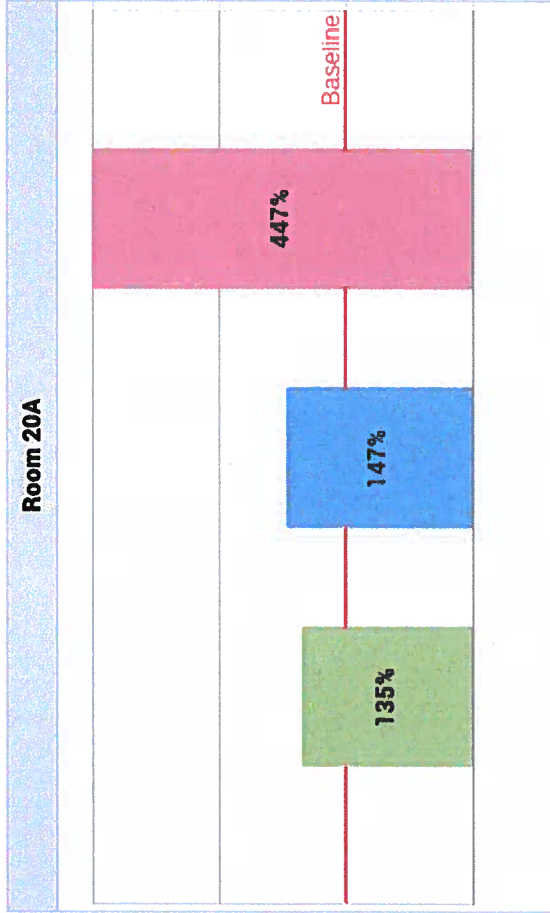
<p>Reporting Limit</p>	<p>The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.</p>
<p>Blanks</p>	<p>Results have not been corrected for field or laboratory blanks.</p>
<p>Background</p>	<p>The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of Aspergillus and Penicillium may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows:</p> <p>NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD)</p> <p>1 : <5% of field occluded. No spores will be uncountable. 2 : 5-25% of field occluded. 3 : 25-75% of field occluded. 4 : 75-90% of field occluded. 5 : >90% of field occluded. Suggested recollection of sample.</p>
<p>Fragments</p>	<p>Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.</p>
<p>Control Comparisons</p>	<p>There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.</p>
<div data-bbox="982 1627 1274 2047" data-label="Figure"> </div> <p>Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.</p> <p>Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.</p> <p>Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.</p> <p>Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.</p> <p>Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.</p>	
<p>Color Coding</p>	<p>Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.</p>

Spore Estimate		Percentages
ND	None Detected	0%
Rare	Less than 10 spores	< 1%
Light	10 - 99 spores	1-10%
Moderate	100 - 999 spores	11-25%
Heavy	1000 - 9999 spores	26-50%
Very Heavy	10000 or greater spores	51-100%

Mycelial Estimate	
ND	None Detected No active growth at site.
Trace	Very small amount of Mycelium Probably no active growth at site.
Few	Some Mycelium Possible active growth at site.
Many	Large amount of Mycelium Probable active growth at site.



Ascospores	Epicoccum	Aspergillus Penicillium	Cladosporium	Curvularia
Myxomycetes	Bipolaris Drechslera			



Ascospores

Habitat: A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
Effects: Health affects are poorly studied, but many are likely to be allergenic.

Aspergillus/Penicillium

Habitat: The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates.
Effects: This group contains common allergens and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.

Bipolaris/Drechslera

Habitat: They are found in soil and as plant pathogens. Can grow indoors on a variety of substrates.
Effects: They may be allergenic and are very commonly involved in allergic fungal sinusitis. They are opportunistic pathogens but occasionally infect healthy individuals, causing keratitis, sinusitis and osteomyelitis.

Cladosporium

Habitat: One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
Effects: A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Curvularia

Habitat: They exist in soil and plant debris, and are plant pathogens.
Effects: They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and disseminated infection, primarily in the immunocompromised.

Epicoccum

Habitat: It is found in soil and plant litter and is a plant pathogen. It can grow indoors on a variety of substrates, including paper and textiles and is commonly found on wet drywall.
Effects: It is a common allergen. No cases of infection have been reported in humans.

Jim McManus
ALIS Environmental Inc.
1027 Koontz Haven Rd
Pinnacle, NC 27043
(336) 368-4500

201-2001-02
Eastlawn Elementary School
502 N. Graham - Hopedale Rd
Burlington, NC

#20004000

Organism Descriptions

Myxomycetes

Habitat: Found on decaying plant material and as a plant pathogen.

Effects: Some allergenic properties reported, but generally pose no health concerns to humans.



Environmental, Inc.
PO Box 6 Pinnacle, NC 27043

Invoice

DATE	INVOICE #
2/2/2020	1766

BILL TO
Alamance-Burlington School System Jay Fuller 307 Prison Camp Road Graham, NC 27253

PROJECT ADDRESS
Eastlawn ES 502 N. Graham-Hopedale Rd Burlington, NC
CLIENT PROJECT ID # / P.O.#

PROJECT NAME	PROJECT #	TERMS	DUE DATE	
Room 20A Air Testing	201-2001-02	Due on receipt	2/2/2020	
PROJECT DATE (S)	SERVICE (S)	DAYS/HOURS/EACH	RATE	AMOUNT
1/29/2020	Airborne Mold Spore Testing	1	425.00	425.00
1/29/2020	Lab Services	4	50.00	200.00

A late charge of 1.5% (18% APR) will be added each month to all invoices 30 days past due.

Invoice Total \$625.00

Thank you for your business !
Please Remit to: P.O. Box 6 Pinnacle, NC 27043

Customer Total Balance \$625.00



Environmental, Inc.
PO Box 6 Pinnacle, NC 27043

Invoice

DATE	INVOICE #
9/22/2020	1865

BILL TO
Alamance-Burlington School System Accounts Payable 307 Prison Camp Road Graham, NC 27253

PROJECT ADDRESS
E.M. Holt ES 4751 S. NC Hwy 62 Burlington, NC
CLIENT PROJECT ID # / P.O.#

PROJECT NAME	PROJECT #	TERMS	DUE DATE	
Mold Assessment	201-2009-05	Due on receipt	9/22/2020	
PROJECT DATE (S)	SERVICE (S)	DAYS/HOURS/EACH	RATE	AMOUNT
9/18/2020	Airborne Fungal Sampling	1	425.00	425.00
9/18/2020	Lab Charges/Sample Analysis	8	50.00	400.00
A late charge of 1.5% (18% APR) will be added each month to all invoices 30 days past due.			Invoice Total	\$825.00

Thank you for your business !
Please Remit to: P.O. Box 6 Pinnacle, NC 27043

Customer Total Balance **\$825.00**



Office: 336.368.4500
Mobile: 336.575.2343
jmcmanus@alisenvironmental.com

September 21, 2020

Jay Fuller
Director of Facilities and Maintenance

Alamance-Burlington School System
307 Prison Camp Road
Graham, North Carolina 27253

Subject: Report of Airborne Microbial Sampling
E M Holt Elementary School – K100 Wing
4751 S. NC Hwy 62
Burlington, North Carolina
Project No.: 201-2009-05

Dear Mr. Fuller:

ALIS has completed the airborne microbial sampling at E M Holt Elementary School in Burlington, North Carolina. On September 18, 2020, at your request, we conducted air sampling for the presence of airborne fungi at the subject property. The purpose of our sampling was to determine the presence and species of airborne fungi and the degree of concentration within the area of concern. One air sampling pump was calibrated and placed inside each of the following classrooms: 101, 102, 107, 108, 111, 112. One sample was collected from outside the building as a reference (baseline) for comparison to the inside conditions. Temperature and relative humidity readings were obtained from each of the tested areas. No other areas of the facility were included in the scope of work.

Results

The laboratory results found the total spore counts on the indoor samples to be generally lower when compared to the outdoor sample. Although spores were detected at lower levels indoors, the sample obtained from room 108 exhibited the species Ascospores and Basidiospores at levels greater than was detected in the remaining classrooms. Additionally, the species Bipolaris/Drechslera was found on three indoor only samples with the higher level found in room 108. Bipolaris/Drechslera was not detected on the outdoor sample. The relative humidity reading in room 108 was highest of all the tested areas. Therefore, we suspect that mold growth may be active in room 108.

Although visible growth was not observed in room 108 during our visit, the heating and air conditioning system should be inspected for condensation, proper drainage and general level of performance-efficiency. Temperature and relative humidity readings obtained inside and outside are as follows:

<u>Rm. 101:</u>	<u>Rm. 102</u>	<u>Rm. 107</u>	<u>Rm. 108</u>	<u>Rm. 111</u>	<u>Rm. 112</u>	<u>Outside</u>
• T: 72.3 ⁰	T: 72.5 ⁰	T: 72.1 ⁰	T: 71.9 ⁰	T: 72.5 ⁰	T: 71.7 ⁰	T: 79.5 ⁰
• RH: 60.6%	RH: 65.6%	RH: 63.3%	RH: 69.8%	RH: 63.3%	RH: 57.5%	RH: 59.5%

Sample results and additional information on fungal spores are attached to this report: “Spore Trap Analysis”

Sampling Methodology

Non-viable samples were collected with a spore trap slide using Allergenco-D Cassettes mounted to a sampling pump. The cassettes contain glass slides that are coated with a sticky substance that captures airborne particulates that impinge on the slides. The air samples were collected at 15 liters per minute for 10 minutes. Calibration of sampling equipment was performed with a precision rotameter (a secondary calibration source). Rotameters are calibrated against a primary standard. Field calibration was performed before and after sampling. The air samples were sealed for transport to Hayes Microbial Consulting in Midlothian, Virginia for analysis. Hayes Microbial is a participant in the American Industrial Hygiene Association, Laboratory Accreditation Program (AIHA-LAP) for Environmental Microbiology.

Background Information on Mold in Buildings

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ALIS appreciates the opportunity to be of service to you on this project. We would welcome the opportunity to discuss at your convenience, any of the results contained in this report. Please contact us if you have any questions or if we may be of further service.

Sincerely,
ALIS ENVIRONMENTAL, INC.



James P. McManus
Vice-President

Attachment: “Spore Trap Analysis”



HAYES
MICROBIAL CONSULTING

Chain of N
3005 East Bounded,
Midlothian,
Ph. 804.562.3435 F

SHIP: FEDEX - PAK 50
DATE: 09-21-2020

8054 0522 9192

MOLD



200333558

ALIS Environmental, Inc.
1027 Koontz Haven Rd
Pinnacle, NC 27043
Ph.: 336.575.2343 Fax:

Job Number: 201-2009-05
Job Name:
E.M. Holt Elementary School
4751 S. NC Hwy. 62
Burlington, NC

Date Collected: 9/18/2020
Collected by: J. McManus
Email: jmcmanus@alisenvironmental.com

Sample #	Sample Name	Analysis Type	Volume	Turn Around Time	Start / Stop Time
5387	Rm 112	S	150 LTR	24 HRS	12:55 / 13:05 T: 71.7 RH: 57.5
6123	"	S	165		13:00 / 13:11 T: 72.5 RH: 63.3
9067	"	S	165		13:09 / 13:20 T: 71.9 RH: 69.8
6859	"	S	165		13:15 / 13:26 T: 73.1 RH: 68.3
8883	"	S	150		13:24 / 13:34 T: 72.5 RH: 65.6
6022	"	S	150		13:30 / 13:40 T: 72.3 RH: 69.3
4370	"	S	150		13:52 / 14:02 T: 74.5 RH: 69.5
6290	BLANK	S	-		-

Analysis Type	Description	Turn Around Time	Acceptable Samples Types
Spore Trap	S Identification & Enumeration of Fungal Spores	24 hours	Spore Trap cassettes, Impact slides
	S+ f & E of Fungal Spores + total dander, fiber and pollen count	24 hours	Spore Trap cassettes, Impact slides
Direct ID	D ID and Semi-quantitative enumeration of spores and mycelium	24 hours	Tape, Bio-tape, swab, bulk, agar plate for ID only
	D+ ID and Enumeration with spore count	24 hours	Tape, Bio-tape, swab, bulk, agar plate for ID only
Culture	C1 Identification & Enumeration of Mold only	7 days	Anderson Air Plate, Swab, Bulk
	C2 Identification & Enumeration of Bacteria only	4 days	Anderson Air Plate, Swab, Bulk
	C3 Identification & Enumeration of Mold and Bacteria	7 days	Anderson Air Plate, Swab, Bulk
Dust Mite	A1 Semi-quantitative analysis of dust mite allergen	24 hours	Bulk Dust

Notes:

Relinquished By: *J. McManus* Date: 9-18-20 Rcvd. By: *CP* Date: 9/21/20 Time:



#20033558

Analysis Report prepared for

ALIS Environmental Inc.

1027 Koontz Haven Rd
Pinnacle, NC 27043

Phone: (336) 368-4500

201-2009-05
E.M. Holt Elementary School
4751 S. NC Hwy 62
Burlington, NC 27215

Collected: **September 18, 2020**
Received: **September 21, 2020**
Reported: **September 21, 2020**



EPA Laboratory ID: VA01419

We would like to thank you for trusting Hayes Microbial for your analytical needs!

We received 8 samples by FedEx in good condition for this project on September 21st, 2020.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.

Steve Hayes, BSMT(ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.



Lab ID: #188863



DPH License: #PH-0198

Sample Number	5387	6123	9067	6859		
Sample Name	Rm 112	Rm 111	Rm 108	Rm 107		
Sample Volume	150.00 liter	165.00 liter	165.00 liter	165.00 liter		
Reporting Limit	7 spores/m ³	6 spores/m ³	6 spores/m ³	6 spores/m ³		
Background	2	2	2	2		
Fragments	ND	18/m ³	ND	ND		
Organism	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total
Alternaria						
Asco spores	2	13	66.7%	8	48	72.7%
Aspergillus Penicillium						
Basidiospores	1	7	33.3%	2	12	18.2%
Bipolaris Drechslera						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Memnoniella						
Myxomycetes						
Pithomyces						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Cercospora						
Total	3	20	100%	11	66	100%

Water Damage Indicator Common Allergen Slightly Higher than Baseline Significantly Higher than Baseline Ratio Abnormality

Collected: Sep 18, 2020 Received: Sep 21, 2020 Reported: Sep 21, 2020
 Project Analyst: Ramesh Poluri, PhD P. Ramesh Date: 09-21-2020
 Reviewed By: Steve Hayes, BSMT Stephen A. Hayes Date: 09-21-2020
 contact@hayesmicrobial.com (804) 562-3435

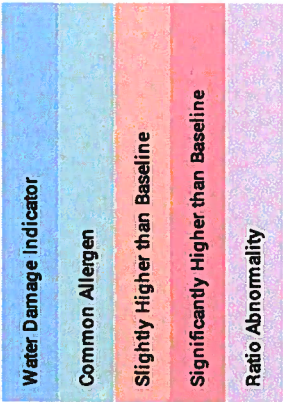


Sample Number	8883	6022	4370	6390		
Sample Name	Rm 102	Rm 101	Rm O/S	Blank		
Sample Volume	150.00 liter	150.00 liter	150.00 liter	0.00 liter		
Reporting Limit	7 spores/m ³	7 spores/m ³	7 spores/m ³	1 spore/m ³		
Background	2	2	2	NBD		
Fragments	7/m ³	ND	ND	ND		
Organism	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total
Alternaria						
Ascospores	4	27	19.0%	360	2400	54.5%
Aspergillus Penicillium				4	27	<1%
Basidiospores	1	7	4.8%	256	1707	38.7%
Bipolaris Drechslera	1	7	4.8%			
Chaetomium						
Cladosporium	7	47	33.3%	32	213	4.8%
Curvularia	5	33	23.8%	3	20	<1%
Epicoccum						
Fusarium						
Memnoniella						
Myxomycetes						
Pithomyces	3	20	14.3%	2	13	<1%
Stachybotrys						
Stemphylium						
Torula				1	7	<1%
Ulocladium						
Cercospora				3	20	<1%
Total	21	141	100%	661	4407	100%

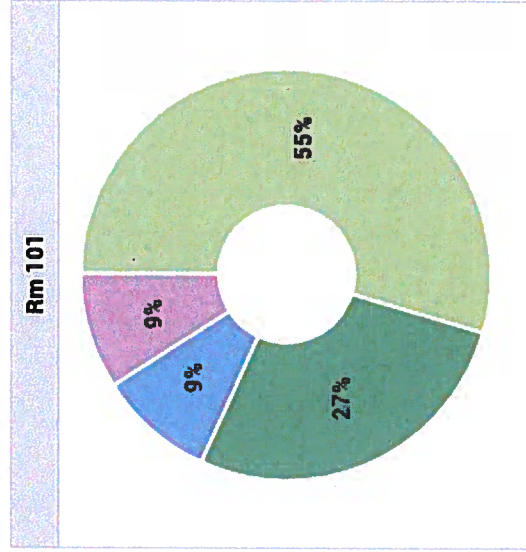
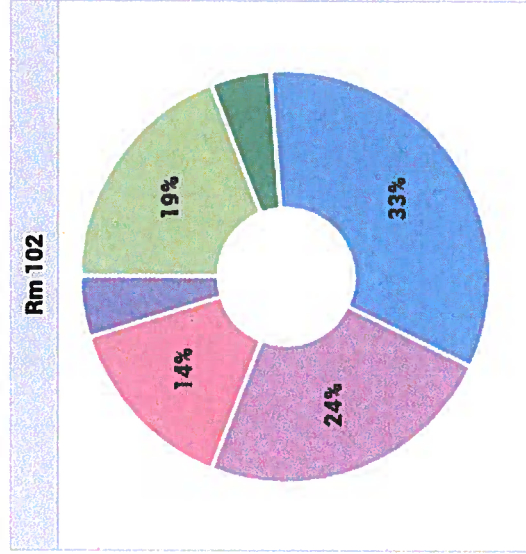
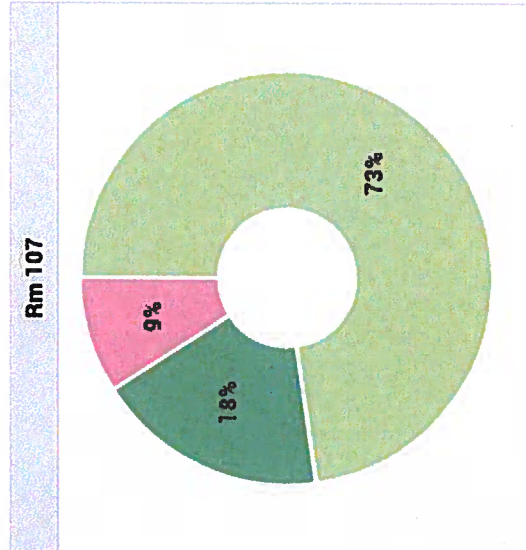
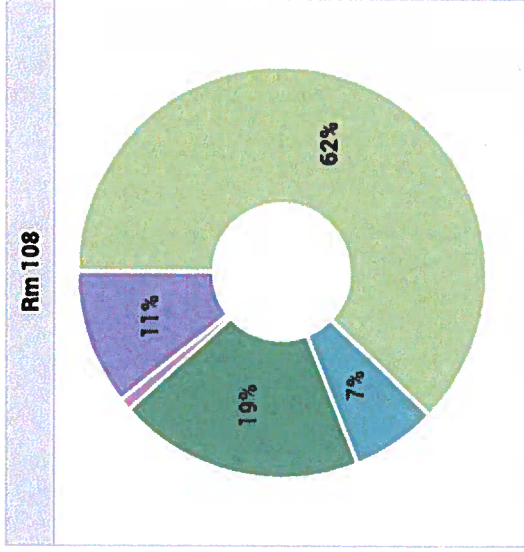
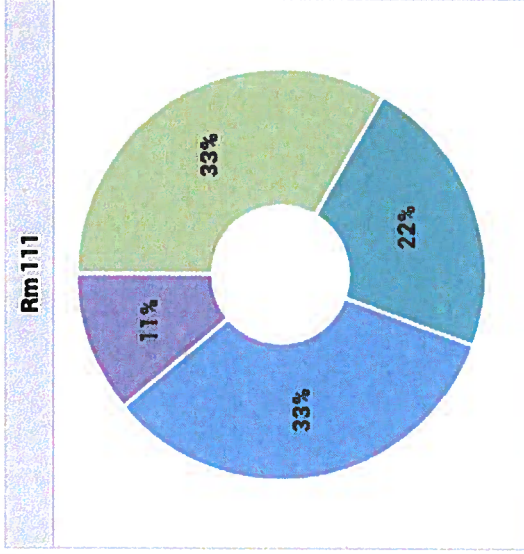
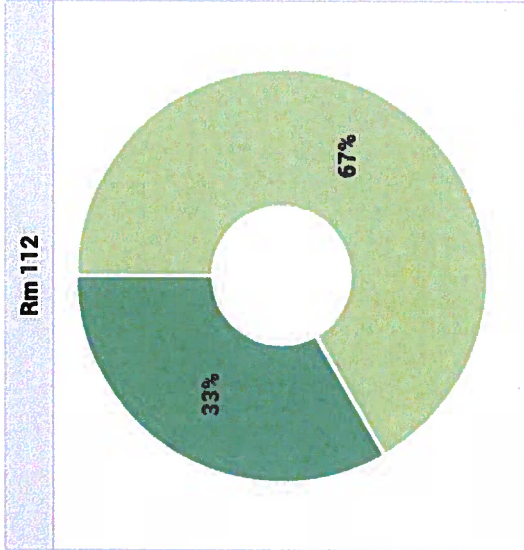
Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
Collected: Sep 18, 2020	Received: Sep 21, 2020	Reported: Sep 21, 2020		
Project Analyst: Ramesh Poluri, PhD	<i>P. Ramesh</i>	Reviewed By: Steve Hayes, BSMT	<i>Stephen A. Hayes</i>	Date: 09 - 21 - 2020
3005 East Boundary Terrace, Suite F. Midlothian, VA. 23112		(804) 562-3435		contact@hayesmicrobial.com



Spore Trap Information

<p>Reporting Limit</p> <p>The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.</p>
<p>Blanks</p> <p>Results have not been corrected for field or laboratory blanks.</p>
<p>Background</p> <p>The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of Aspergillus and Penicillium may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows:</p> <p>NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD)</p> <p>1 : <5% of field occluded. No spores will be uncountable. 2 : 5-25% of field occluded. 3 : 25-75% of field occluded. 4 : 75-90% of field occluded. 5 : >90% of field occluded. Suggested recollection of sample.</p>
<p>Fragments</p> <p>Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.</p>
<p>Control Comparisons</p> <p>There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.</p>
 <p>Water Damage Indicator (Blue)</p> <p>Common Allergen (Light Blue)</p> <p>Slightly Higher than Baseline (Light Green)</p> <p>Significantly Higher than Baseline (Green)</p> <p>Ratio Abnormality (Red)</p> <p>Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.</p> <p>Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.</p> <p>Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.</p> <p>Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.</p> <p>Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.</p>
<p>Color Coding</p> <p>Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.</p>

Spore Counts - Pie Charts

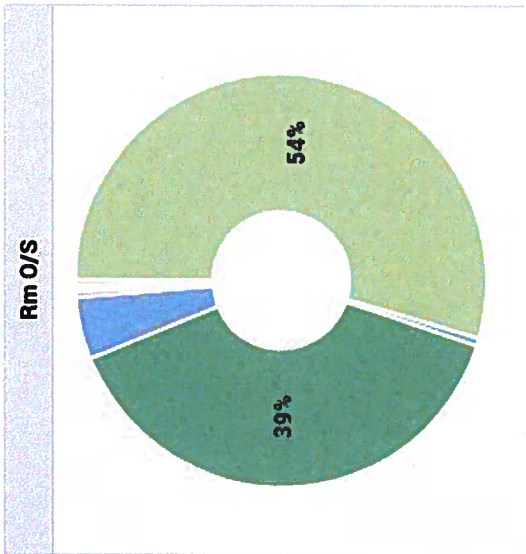


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 4751 S. NC Hwy 62
 Burlington, NC 27215

#20033558

Spore Counts - Pie Charts



Ascospores	Basidiospores	Aspergillus Penicillium	Cladosporium	Bipolaris Drechslera
Curvularia	Pithomyces	Torula	Cercospora	

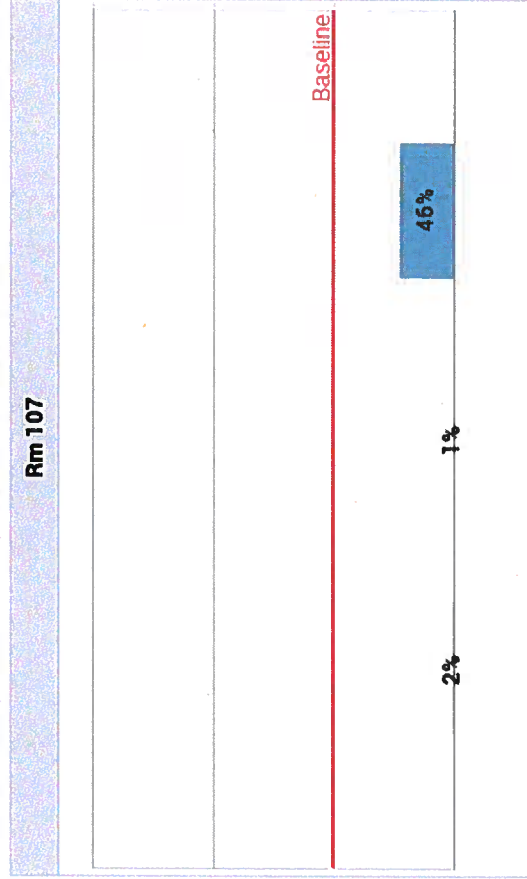
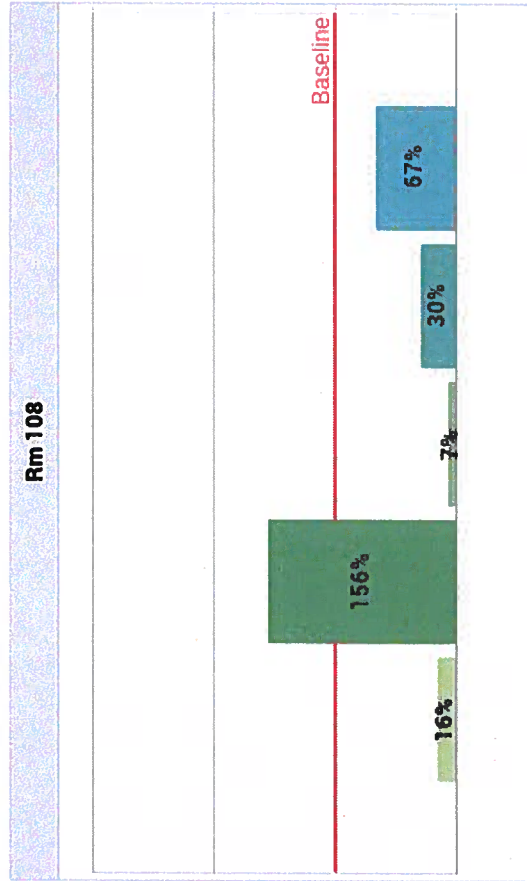
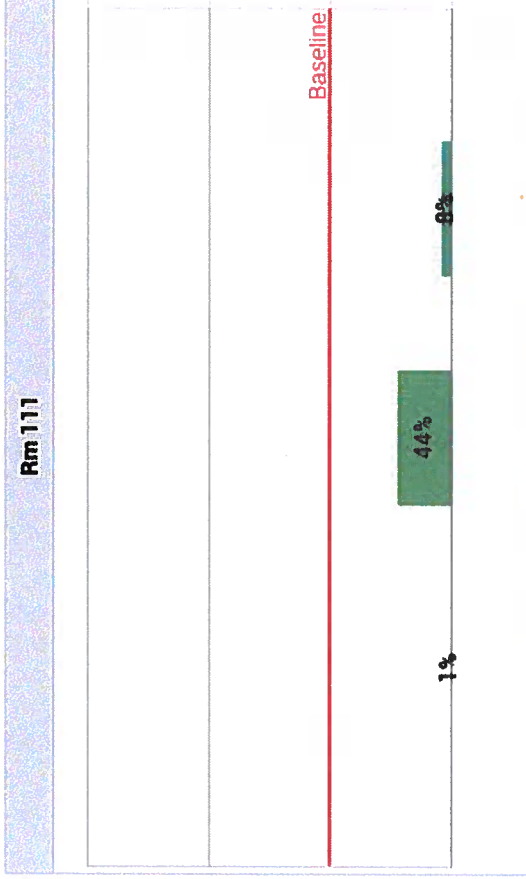
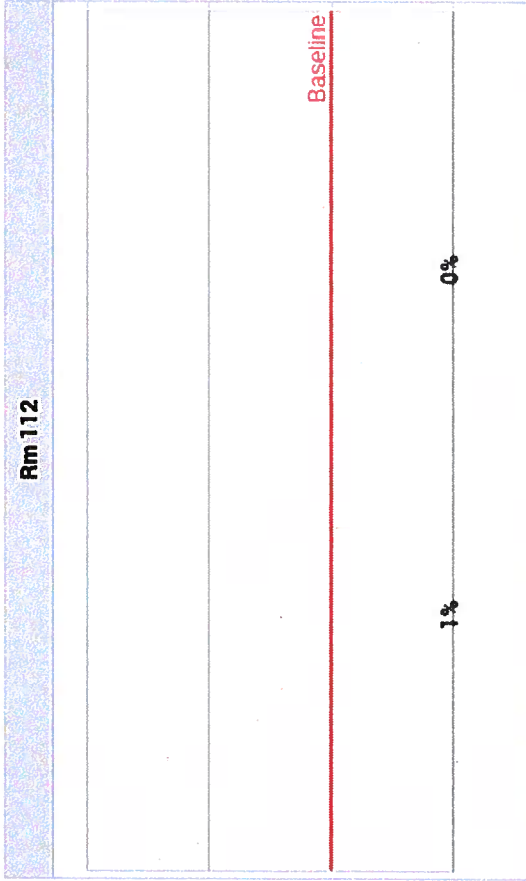


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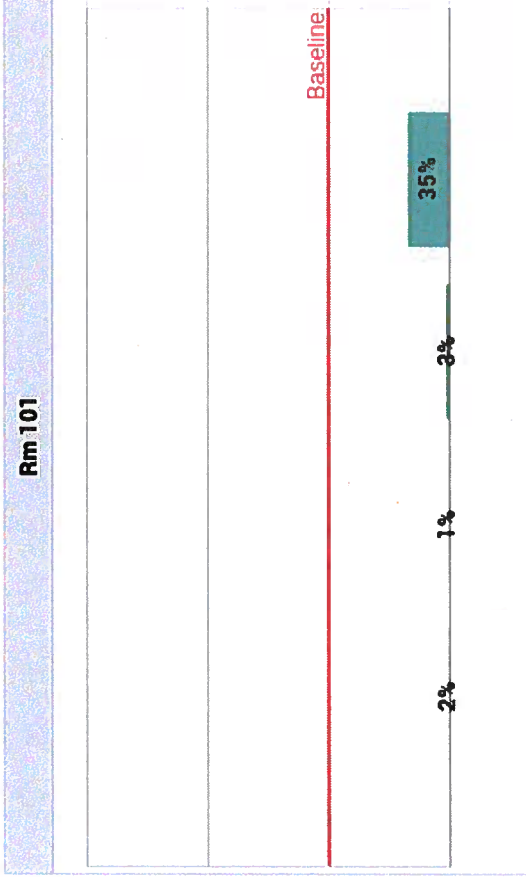
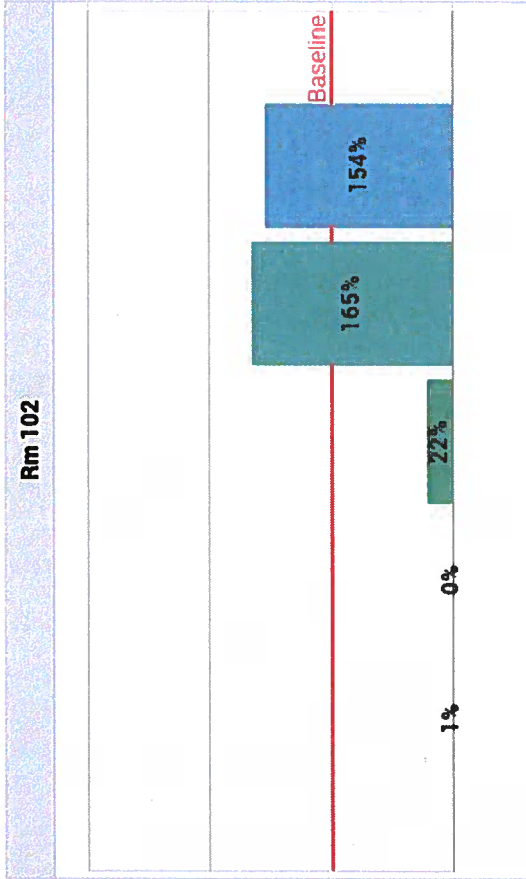
contact@hayesmicrobial.com

Spore Counts - Bar Graphs



Ascospores	Aspergillus/Penicillium	Cladosporium	Curvularia
Bipolaris/Drechslera			

Spore Counts - Bar Graphs



Ascospores	Basidiospores	Aspergillus/Penicillium	Cladosporium	Curvularia
Bipolaris/Drechslera	Pithomyces			

Organism Descriptions

Ascospores

Habitat: A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.
Effects: Health affects are poorly studied, but many are likely to be allergenic.

Aspergillus/Penicillium

Habitat: The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates.
Effects: This group contains common allergens and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.

Basidiospores

Habitat: A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.
Effects: Common allergens and are also associated with hypersensitivity pneumonitis.

Bipolaris/Drechslera

Habitat: They are found in soil and as plant pathogens. Can grow indoors on a variety of substrates.
Effects: They may be allergenic and are very commonly involved in allergic fungal sinusitis. They are opportunistic pathogens but occasionally infect healthy individuals, causing keratitis, sinusitis and osteomyelitis.

Cercospora

Habitat: Found on wood and decaying plant matter.
Effects: Health effects are poorly studied.

Cladosporium

Habitat: One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.
Effects: A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Curvularia

Habitat: They exist in soil and plant debris, and are plant pathogens.

Effects: They are allergenic and a common cause of allergic fungal sinusitis. An occasional cause of human infection, including keratitis, sinusitis, onychomycosis, mycetoma, pneumonia, endocarditis and disseminated infection, primarily in the immunocompromised.

Pithomyces

Habitat: Common fungus isolated from soil, decaying plant material. Rarely found indoors.

Effects: Allergenic properties are poorly studied. No cases of infection in humans.

Torula

Habitat: Found in soil and on wood and grasses. Occasionally found growing indoors on cellulose containing materials.

Effects: A known allergen. No known cases of human infection.



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jmcmanus@allsenvironmental.com

October 16, 2020

Jay Fuller
Director of Facilities and Maintenance

Alamance-Burlington School System
307 Prison Camp Road
Graham, North Carolina 27253

Subject: Report of Airborne Microbial Sampling
Eastern Alamance High School – A Wing
4040 Mebane-Rodgers Road
Mebane, North Carolina
Project No.: 201-2010-06

Dear Mr. Fuller:

ALIS has completed the airborne microbial sampling at Eastern Alamance High School in Mebane, North Carolina. On October 12 2020, at your request, we conducted air sampling for the presence of airborne fungi at the subject property. The purpose of our sampling was to determine the presence and species of airborne fungi and the degree of concentration within the area of concern. One air sampling pump was calibrated and placed inside each of the following classrooms: A1, A2, A3, A4, A5, A6 A8. One sample was collected from outside the building as a reference (baseline) for comparison to the inside conditions. Temperature and relative humidity readings were obtained from each of the tested areas. No other areas of the facility were included in the scope of work.

Results

The laboratory results found the total spore counts on the indoor samples to be generally lower when compared to the outdoor sample. Although spores were detected at lower levels indoors, three spores of the species *Cladosporium* was detected in room A1, and one spore of the species *Pithomyces* was detected in room A4. *Cladosporium* and *Pithomyces* were not detected on the outdoor sample. Although these species were not detected on the outdoor sample, their indoor levels are not sufficient to suspect that active growth is occurring. Visible mold was not observed during our testing. Temperature and relative humidity readings obtained inside and outside are as follows:

<u>Rm. A1</u>	<u>Rm. A2</u>	<u>Rm. A3</u>	<u>Rm. A4</u>	<u>Rm. A5</u>	<u>Rm. A6</u>	<u>Rm. A8</u>
• T: 72.6 ⁰	T: 71.4 ⁰	T: 72.8 ⁰	T: 71.6 ⁰	T: 72.1 ⁰	T: 71.2 ⁰	T: 70.3 ⁰
• RH: 60.1%	RH: 60.3%	RH: 62.9%	RH: 55.3%	RH: 57.3%	RH: 50.6%	RH: 56.0%
 <u>Outdoor</u>						
• T: 71.7 ⁰						
• RH: 78.3%						

Sample results and additional information on fungal spores are attached to this report: "Spore Trap Analysis"

Sampling Methodology

Non-viable samples were collected with a spore trap slide using Allergenco-D Cassettes mounted to a sampling pump. The cassettes contain glass slides that are coated with a sticky substance that captures airborne particulates that impinge on the slides. The air samples were collected at 15 liters per minute for 10 minutes. Calibration of sampling equipment was performed with a precision rotameter (a secondary calibration source). Rotameters are calibrated against a primary standard. Field calibration was performed before and after sampling. The air samples were sealed for transport to Hayes Microbial Consulting in Midlothian, Virginia for analysis. Hayes Microbial is a participant in the American Industrial Hygiene Association, Laboratory Accreditation Program (AIHA-LAP) for Environmental Microbiology.

Background Information on Mold in Buildings

Mold spores exist normally in outdoor and indoor air and can be measured in air and carpets of normal homes, office buildings, hospitals and schools. Naturally occurring sources of mold spores include soil, plants and other sources. The air concentration of these normally occurring mold spores is dependent on the season, environmental conditions and other factors. Elevated levels of mold in building materials may occur if chronic moist conditions from water leaks, floods, chronic high relative humidity, or malfunctioning heating, ventilation or air conditioning systems, allow moisture to remain for prolonged periods on organic matter in the presence of warm ambient temperatures. Under these conditions, low levels of fungal spores in air, plants or other sources, may proliferate on cellulose containing materials such as carpets, wallboard, wood, paper or dusty surfaces (which may serve as a food source), and result in mold contamination. Many fungal spores are allergenic to susceptible persons exposed, though individual susceptibility varies greatly. There is no practical way to eliminate all mold and mold spores in the indoor environment; the way to control indoor mold growth is to control moisture.

ALIS appreciates the opportunity to be of service to you on this project. We would welcome the opportunity to discuss at your convenience, any of the results contained in this report. Please contact us if you have any questions or if we may be of further service.

Sincerely,
ALIS ENVIRONMENTAL, INC.



James P. McManus
Vice-President

Attachment: "Spore Trap Analysis"



#20037243

Analysis Report prepared for

ALIS Environmental Inc.

1027 Koontz Haven Rd
Pinnacle, NC 27043

Phone: (336) 368-4500

201-2010-06
Eastern Alamance High School
4040 Mebane-Rodgers Road
Mebane, NC 27302

Collected: **October 12, 2020**
Received: **October 13, 2020**
Reported: **October 13, 2020**



EPA Laboratory ID: VA01419

We would like to thank you for trusting Hayes Microbial for your analytical needs!
We received 9 samples by FedEx in good condition for this project on October 13th, 2020.

The results in this analysis pertain only to this job, collected on the stated date, and should not be used in the interpretation of any other job. This report may not be duplicated, except in full, without the written consent of Hayes Microbial Consulting, LLC.

This laboratory bears no responsibility for sample collection activities, analytical method limitations, or your use of the test results. Interpretation and use of test results are your responsibility. Any reference to health effects or interpretation of mold levels is strictly the opinion of Hayes Microbial. In no event, shall Hayes Microbial or any of its employees be liable for lost profits or any special, incidental or consequential damages arising out of the use of these test results.

Steve Hayes, BSMT(ASCP)
Laboratory Director
Hayes Microbial Consulting, LLC.



Lab ID: #188863



DPH License: #PH-0198

Sample Number	5755	7043	7411	5654		
Sample Name	Classroom A1	Classroom A2	Classroom A4	Classroom A8		
Sample Volume	150.00 liter	150.00 liter	150.00 liter	150.00 liter		
Reporting Limit	7 spores/m ³	7 spores/m ³	7 spores/m ³	7 spores/m ³		
Background	3	2	2	2		
Fragments	ND	ND	ND	ND		
Organism	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total
Alternaria						
Ascospores	1	7	25.0%	1	7	50.0%
Aspergillus Penicillium						
Basidiospores						
Bipolaris Drechslera						
Chaetomium						
Cladosporium	3	20	75.0%	1	7	50.0%
Curvularia						
Epicoccum						
Fusarium						
Memnoniella						
Myxomycetes						
Pithomyces				1	7	50.0%
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Total	4	27	100%	2	14	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
Collected: Oct 12, 2020	Received: Oct 13, 2020	Reported: Oct 13, 2020		
Project Analyst: Carlie Hampton, BS	Date: 10 - 13 - 2020	Reviewed By: Steve Hayes, BSMT	Date: 10 - 13 - 2020	Signature: <i>Stephen N. Hayes</i>
3005 East Boundary Terrace, Suite F. Midlothian, VA. 23112		(804) 562-3435		contact@hayesmicrobial.com



Sample Number	8699	8515	5203	9435		
Sample Name	Classroom A6	Classroom A5	Classroom A3	Classroom O/S (outside) Jmc		
Sample Volume	150.00 liter	150.00 liter	150.00 liter	150.00 liter		
Reporting Limit	7 spores/m ³	7 spores/m ³	7 spores/m ³	7 spores/m ³		
Background	2	2	2	2		
Fragments	ND	ND	ND	ND		
Organism	Raw Count	Count / m ³	% of Total	Raw Count	Count / m ³	% of Total
Alternaria						
Ascomycetes						
Aspergillus Penicillium	1	7	50.0%	2	13	100.0%
Basidiospores	1	7	50.0%			
Bipolaris Drechslera						
Chaetomium						
Cladosporium						
Curvularia						
Epicoccum						
Fusarium						
Memnoniella						
Myxomycetes						
Pithomyces						
Stachybotrys						
Stemphylium						
Torula						
Ulocladium						
Total	2	14	100%	289	1926	100%

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
Collected: Oct 12, 2020	Received: Oct 13, 2020	Reported: Oct 13, 2020		
Project Analyst: Carlie Hampton, BS	Reviewed By: Steve Hayes, BSMT	Date: 10 - 13 - 2020	Date: 10 - 13 - 2020	Date: 10 - 13 - 2020
3005 East Boundary Terrace, Suite F. Midlothian, VA. 23112	(804) 562-3435	contact@hayesmicrobial.com		Page: 3 of 10




Stephen N. Hayes

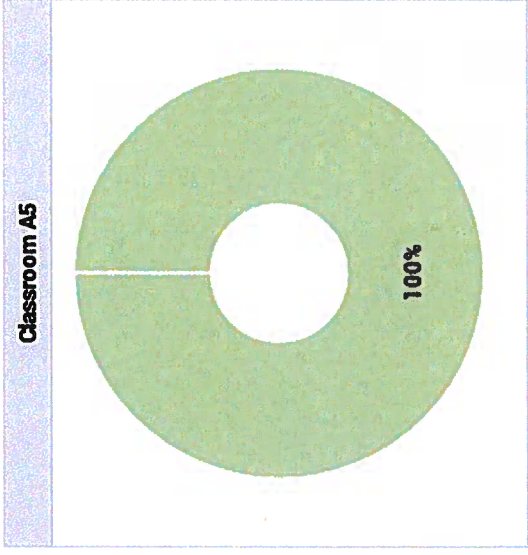
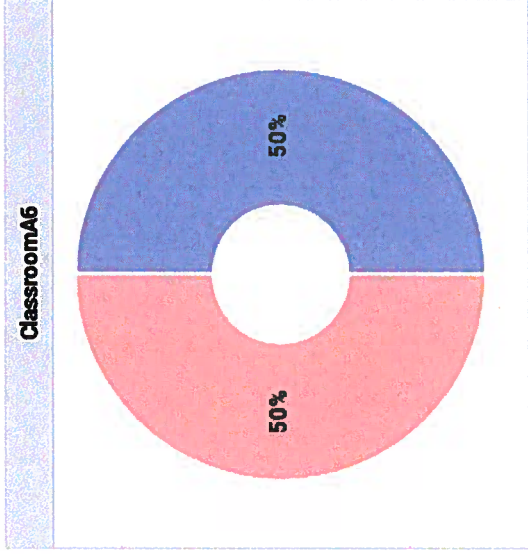
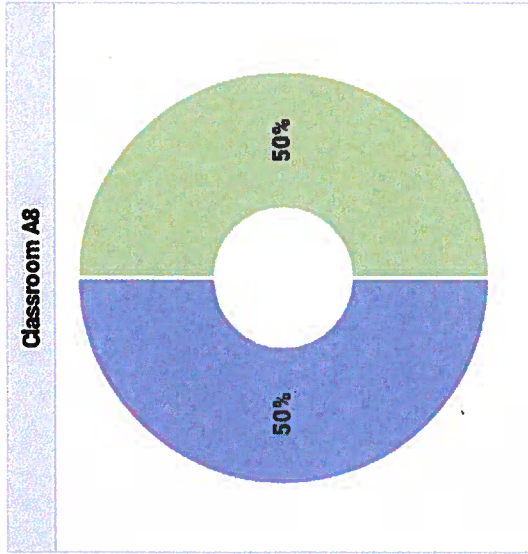
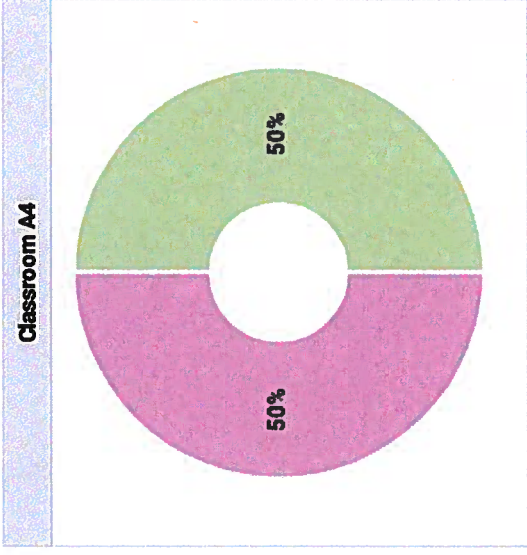
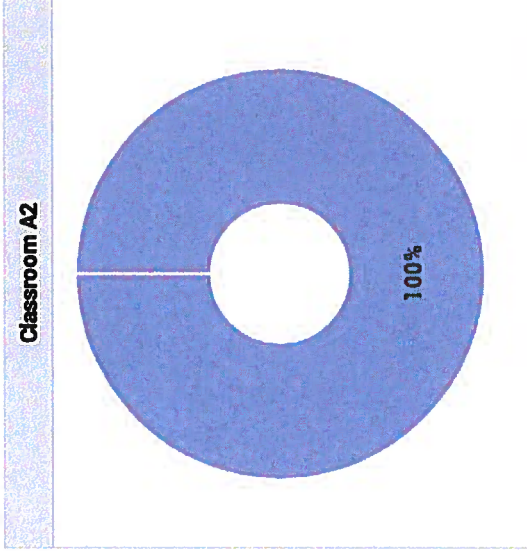
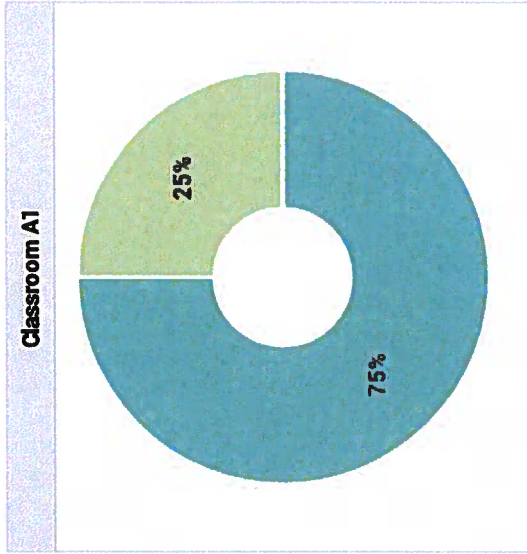
Sample Number	6675			
Sample Name	Classroom B1			
Sample Volume	150.00 liter			
Reporting Limit	7 spores/m ³			
Background	1			
Fragments	ND			
Organism	Raw Count	Count / m ³	% of Total	
Alternaria				
Aspores	1	7	100.0%	
Aspergillus Penicillium				
Basidiospores				
Bipolaris Drechslera				
Chaetomium				
Cladosporium				
Curvularia				
Epicoccum				
Fusarium				
Memnoniella				
Myxomycetes				
Pithomyces				
Stachybotrys				
Stemphylium				
Torula				
Ulocladium				
Total	1	7	100%	

Water Damage Indicator	Common Allergen	Slightly Higher than Baseline	Significantly Higher than Baseline	Ratio Abnormality
Collected: Oct 12, 2020	Received: Oct 13, 2020	Reported: Oct 13, 2020	Reviewed By: Steve Hayes, BSMT	Date: 10-13-2020
Project Analyst: Carlie Hampton, BS	Reviewed By: <i>Stephen N. Hayes</i> contact@hayesmicrobial.com			
3005 East Boundary Terrace, Suite F. Midlothian, VA. 23112 (804) 562-3435				



<p>Reporting Limit</p> <p>The Reporting Limit is the lowest number of spores that can be detected based on the total volume of the sample collected and the percentage of the slide that is counted. At Hayes Microbial, 100% of the slide is read so the LOD is based solely on the total volume. Raw spore counts that exceed 500 spores will be estimated.</p>
<p>Blanks</p> <p>Results have not been corrected for field or laboratory blanks.</p>
<p>Background</p> <p>The Background is the amount of debris that is present in the sample. This debris consists of skin cells, dirt, dust, pollen, drywall dust and other organic and non-organic matter. As the background density increases, the likelihood of spores, especially small spores such as those of Aspergillus and Penicillium may be obscured. The background is rated on a scale of 1 to 5 and each level is determined as follows:</p> <p>NBD: No background detected due to possible pump or cassette malfunction. Recollect sample. (Field Blanks will display NBD)</p> <p>1 : <5% of field occluded. No spores will be uncountable. 2 : 5-25% of field occluded. 3 : 25-75% of field occluded. 4 : 75-90% of field occluded. 5 : >90% of field occluded. Suggested recollection of sample.</p>
<p>Fragments</p> <p>Fragments are small pieces of fungal mycelium or spores. They are not identifiable as to type and when present in very large numbers, may indicate the presence of mold amplification.</p>
<p>Control Comparisons</p> <p>There are no national standards for the numbers of fungal spores that may be present in the indoor environment. As a general rule and guideline that is widely accepted in the indoor air quality field, the numbers and types of spores that are present in the indoor environment should not exceed those that are present outdoors at any given time. There will always be some mold spores present in "normal" indoor environments. The purpose of sampling and counting spores is to help determine whether an abnormal condition exists within the indoor environment and if it does, to help pinpoint the area of contamination. Spore counts should not be used as the sole determining factor of mold contamination. There are many factors that can cause anomalies in the comparison of indoor and outdoor samples due to the dynamic nature of both of those environments.</p>
 <p>Water Damage Indicator (Blue)</p> <p>Common Allergen (Green)</p> <p>Slightly Higher than Baseline (Orange)</p> <p>Significantly Higher than Baseline (Red)</p> <p>Ratio Abnormality (Violet)</p> <p>Blue: These molds are commonly seen in conditions of prolonged water intrusion and usually indicate a problem.</p> <p>Green: Although all molds are potential allergens, these are the most common allergens that may be found indoors.</p> <p>Orange: The spore count is slightly higher than the outside count and may or may not indicate a source of contamination.</p> <p>Red: The spore count is significantly higher than the baseline count and probably indicates a source of contamination.</p> <p>Violet: The types of spores found indoors should be similar to the ones that were identified in the baseline sample. Significant increases (more than 25%) in the ratio of a particular spore type may indicate the presence of abnormal levels of mold, even if the total number of spores of that type is lower in the indoor environment than it was outdoors.</p>
<p>Color Coding</p> <p>Fungi that are present in indoor samples at levels lower than 200 per cubic meter are not color coded on the report, unless they are one of the water damage indicators.</p>

Spore Counts - Pie Charts

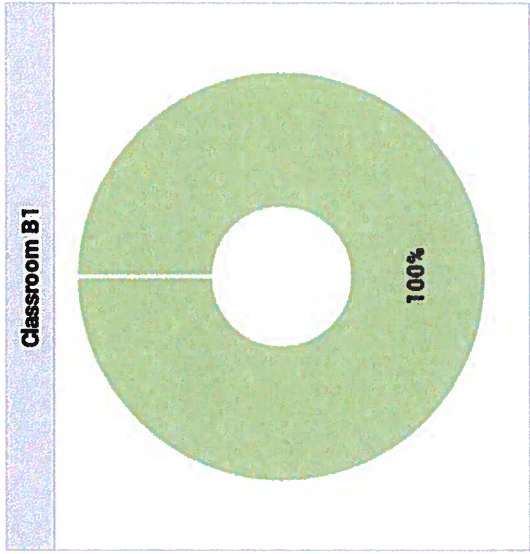
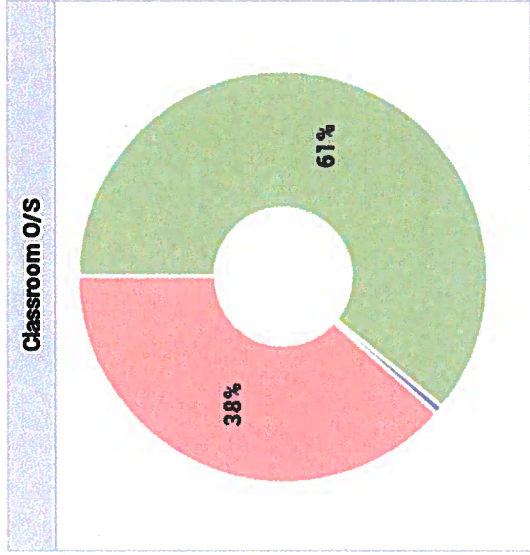
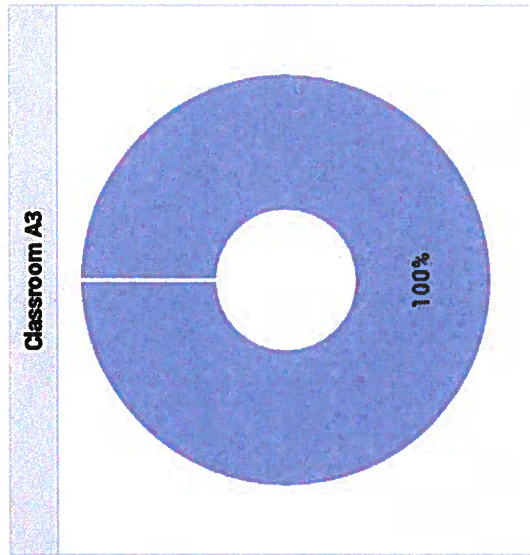


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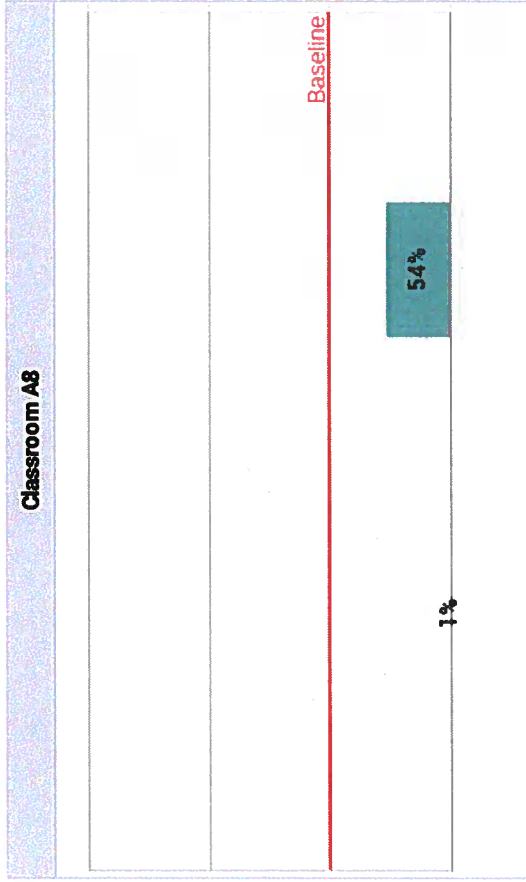
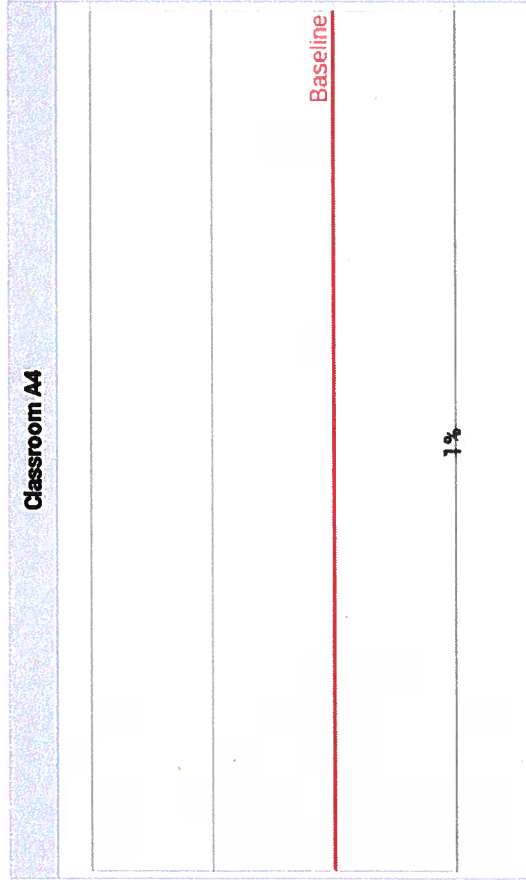
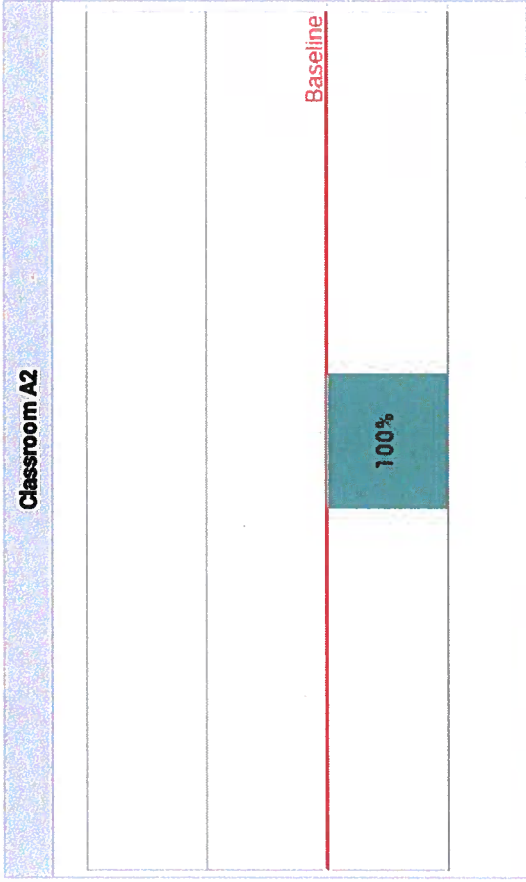
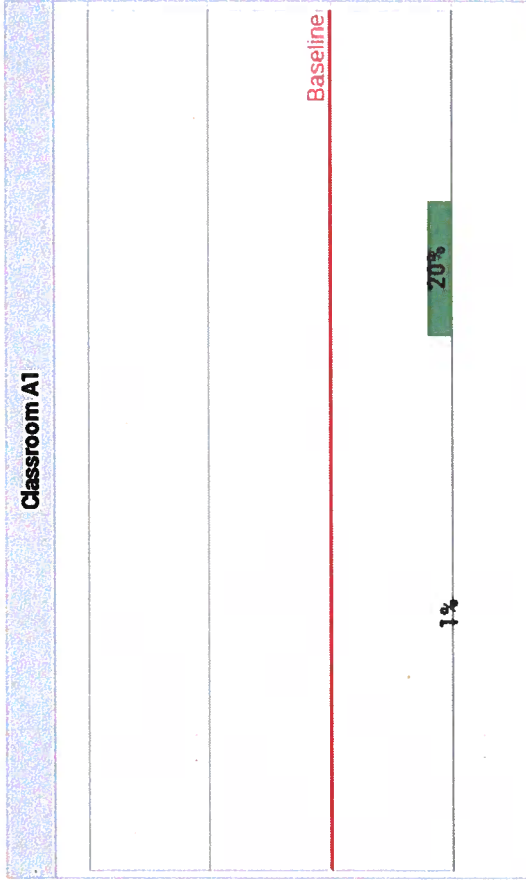
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Spore Counts - Pie Charts

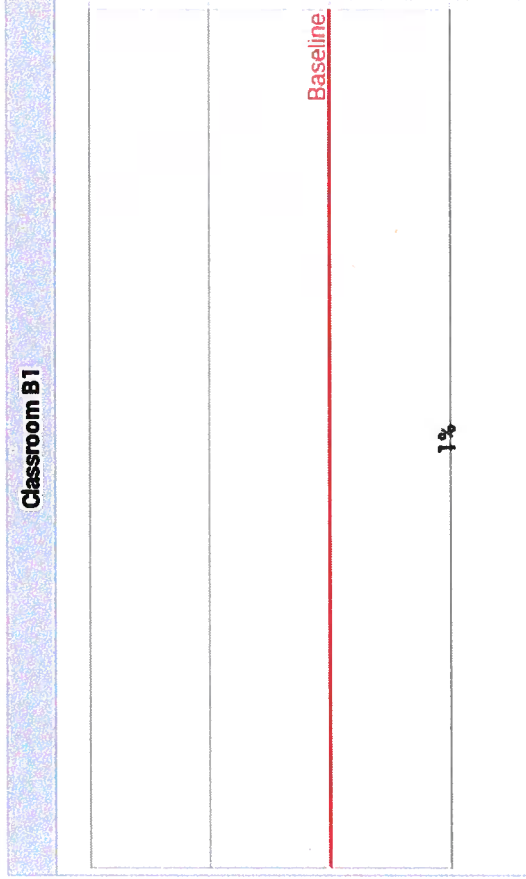
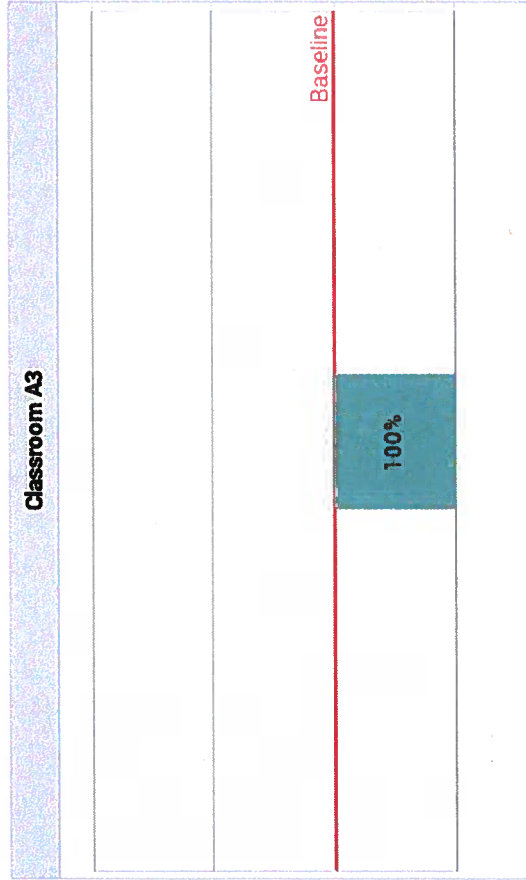
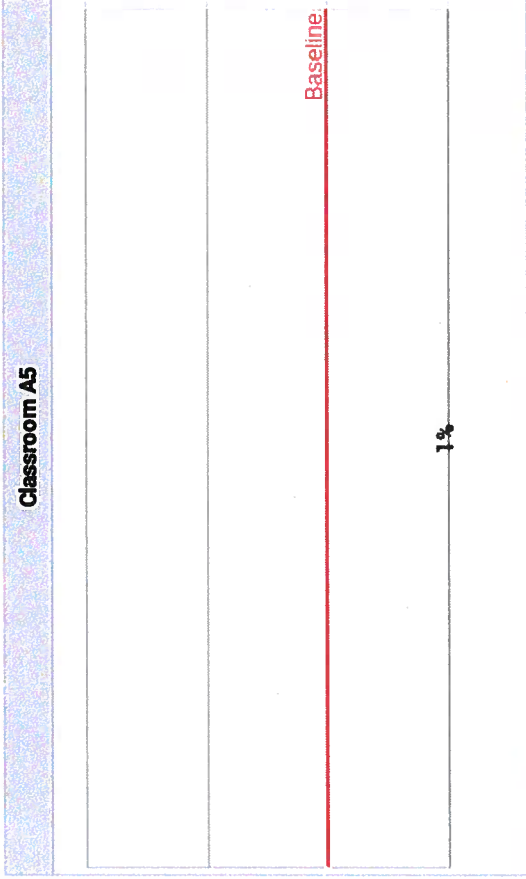
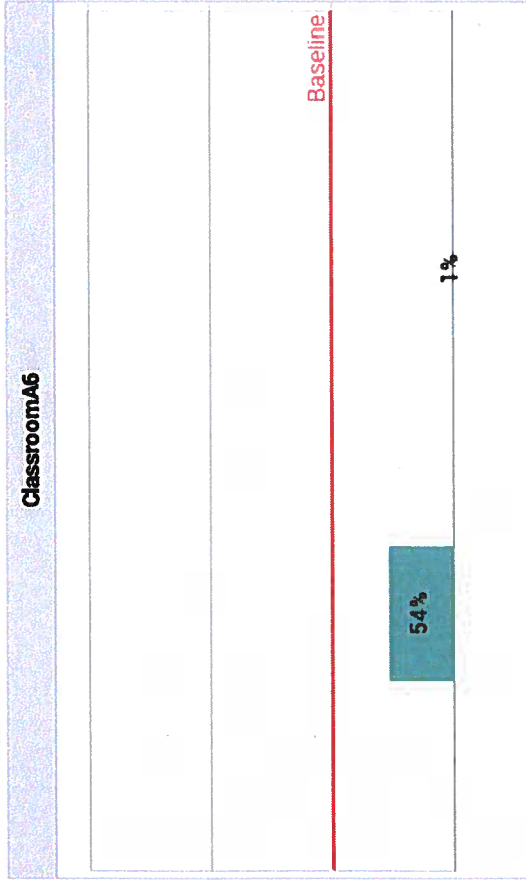


Ascospores	Aspergillus/Penicillium	Pithomyces	Basidiospores
Cladosporium			

Spore Counts - Bar Graphs



Spore Counts - Bar Graphs



Organism Descriptions

Ascospores

Habitat: A large group consisting of more than 3000 species of fungi. Common plant pathogens and outdoor numbers become very high following rain. Most of the genera are indistinguishable by spore trap analysis and are combined on the report.

Effects: Health affects are poorly studied, but many are likely to be allergenic.

Aspergillus/Penicillium

Habitat: The most common fungi isolated from the environment. Very common in soil and on decaying plant material. Are able to grow well indoors on a wide variety of substrates.

Effects: This group contains common allergens and many can cause hypersensitivity pneumonitis. They may cause extrinsic asthma, and many are opportunistic pathogens. Many species produce mycotoxins which may be associated with disease in humans and other animals. Toxin production is dependent on the species, the food source, competition with other organisms, and other environmental conditions.

Basidiospores

Habitat: A common group of Fungi that includes the mushrooms and bracket fungi. They are saprophytes and plant pathogens. In wet conditions they can cause structural damage to buildings.

Effects: Common allergens and are also associated with hypersensitivity pneumonitis.

Cladosporium

Habitat: One of the most common genera worldwide. Found in soil and plant debris and on the leaf surfaces of living plants. The outdoor numbers are lower in the winter and often relatively high in the summer, especially in high humidity. The outdoor numbers often spike in the late afternoon and evening. Indoors, it can be found growing on textiles, wood, sheetrock, moist window sills and in HVAC supply ducts.

Effects: A common allergen, producing more than 10 allergenic antigens and a common cause of hypersensitivity pneumonitis.

Phthomyces

Habitat: Common fungus isolated from soil, decaying plant material. Rarely found indoors.

Effects: Allergenic properties are poorly studied. No cases of infection in humans.



HAYES
MICROBIAL CONSULTING

Chain of Command
3005 East Boundary
Midlothian, VA
Ph. 804.562.3435 Fax. 804-447-0502

SHIP: FEDEX - PAK 50
DATE: 10-13-2020
8054 0522 9207



ALJS Environmental, Inc.
1027 Koontz Haven Rd
Pinnacle, NC 27043
Ph.: 336.575.2343 Fax:

Job Number: 201-2010-06
Job Name: Eastern A;amance High School
4040 Mebane-Rodgers Road
Mebane, NC

Date Collected: 10/12/2020
Collected by: J. McManus
Email: jimcmanus@alisenvironmental.com

Sample #	Sample Name	Analysis Type	Volume	Turn Around Time	Start / Stop Time
5755	CLASSROOM A1	S	150 LTR	24 HR	10:26 / 10:36
7043	A2				10:29 / 10:39
7411	A4				10:33 / 10:43
5654	A8				10:46 / 10:50
8699	A6				10:47 / 10:57
8515	A5				10:58 / 11:02
5203	A3				11:08 / 11:18
9485	o/s				11:26 / 11:36
6625	B1				

Analysis Type	Description	Turn Around Time	Acceptable Samples Types
Spore Trap	S Identification & Enumeration of Fungal Spores	24 hours	Spore Trap cassettes, Impact slides
	S+ I & E of Fungal Spores + total dander, fiber and pollen count	24 hours	Spore Trap cassettes, Impact slides
Direct ID	D ID and Semi-quantitative enumeration of spores and mycellium	24 hours	Tape, Bio-tape, swab, bulk, agar plate for ID only
	D+ ID and Enumeration with spore count	24 hours	Tape, Bio-tape, swab, bulk, agar plate for ID only
Culture	C1 Identification & Enumeration of Mold only	7 days	Anderson Air Plate, Swab, Bulk
	C2 Identification & Enumeration of Bacteria only	4 days	Anderson Air Plate, Swab, Bulk
	C3 Identification & Enumeration of Mold and Bacteria	7 days	Anderson Air Plate, Swab, Bulk
Dust Mite	A1 Semi-quantitative analysis of dust mite allergen	24 hours	Bulk Dust

Notes:

Relinquished By: *C. P. McManus* Date: 10/12/20 Rcvd. By: *JM* Date: 10/13/20 Time:

Handwritten signature/initials

Cloudy / INTERMITTENT Pk

N.O.
o.
N.C.